

SYNTHESIS AND ANTIBACTERIAL
ACTIVITIES OF LESS CHLORINATED
ANALOGS OF NEOPYRROLOMYCIN

KUNIAKI TATSUTA and MANABU ITOH

Graduate School of Science and Engineering,
Waseda University,
3-4-1 Ohkubo, Shinjuku, Tokyo 169, Japan

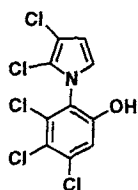
(Received for publication January 25, 1994)

Neopyrrolomycin (**1**) isolated from cultured broth of *Streptomyces* sp. is an optically active and structurally unique phenylpyrrole antibiotic.¹ Very recently, (+)- and (-)-neopyrrolomycins (**1**)² and

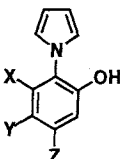
their analogs³ were synthesized from 3,5-dichloroanisole in our laboratories.⁴ Herein we report the synthesis of a variety of less chlorinated analogs (**2**~**14**) and their structure-activity relationship.

All analogs (**2**~**14**) were synthesized from 2-aminoanisoles **15a**~**15d** by our procedures as reported previously.^{2~4} The compound **15a** (oil) was prepared from 3,5-dichlorophenol by nitration with fum. HNO₃ in AcOH successively followed by methylation with Me₂SO₄ to give **24** (mp 72~74°C) and reduction on Pd-C in 34% yield. Compound **15b** (oil) was prepared from **25**⁵ by reduction with Na₂S₂O₄ in 58% yield. Both compounds **15c** and **15d** are commercially available.

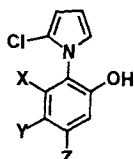
Reaction of **15a**~**15d** with 2,5-dimethoxytetrahy-



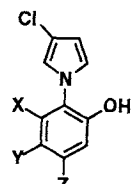
1: Neopyrrolomycin



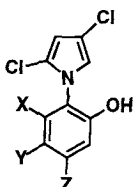
- 2**: X = Z = Cl, Y = H
3: X = Cl, Y = Z = H
4: Y = Cl, X = Z = H
5: X = Y = Z = H



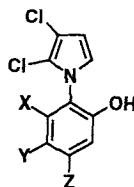
- 6**: X = Z = Cl, Y = H
7: X = Cl, Y = Z = H



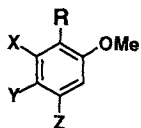
- 8**: X = Cl, Y = Z = H
9: X = Y = Z = H



- 10**: X = Cl, Y = Z = H
11: Y = Cl, X = Z = H
12: X = Y = Z = H



- 13**: X = Z = Cl, Y = H
14: X = Cl, Y = Z = H



- a**: X = Z = Cl, Y = H
b: X = Cl, Y = Z = H
c: Y = Cl, X = Z = H
d: X = Y = Z = H

- 15**: R = NH₂

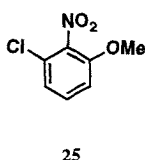
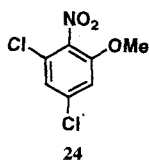
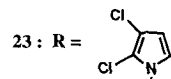
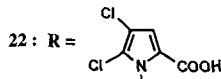
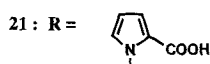
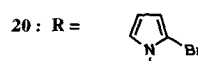
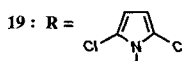
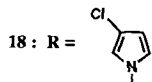
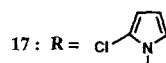
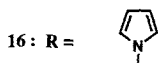


Table 1. Antibacterial and antifungal activities of neopyrrolomycin analogs.^a

Test organism	MIC ($\mu\text{g/ml}$)						
	1 ^b	2	3	4	5	6	7
<i>Citrobacter freundii</i> IFO 12681	50	>50	>50	>100	>100	>50	>50
<i>Enterobacter cloacae</i> IFO 12935	25	1.56	>50	100	100	12.5	>50
<i>Escherichia coli</i> NIHJ JC-2	50	1.56	>50	100	100	12.5	>50
<i>Klebsiella pneumoniae</i> IFO 3317	12.5	0.10	12.5	25	25	0.78	25
<i>Proteus vulgaris</i> GN 5298	25	0.78	>50	100	100	3.13	>50
<i>Pseudomonas aeruginosa</i> IFO 3445	50	>50	>50	>100	>100	>50	>50
<i>Serratia marcescens</i> 3759	50	>50	>50	100	>100	>50	>50
<i>Streptococcus epidermidis</i> IFO 13889	0.39	3.13	>50	100	>100	1.56	50
<i>Enterococcus faecalis</i> IFO 12964	0.78	25	>50	100	>100	3.13	>50
<i>E. faecium</i> IFO 12367	0.20	25	>50	50	>100	1.56	50
<i>Staphylococcus aureus</i> IFO 12732	0.20	0.78	>50	50	>100	1.56	50
Methicillin-resistant <i>S. aureus</i> 4 ^c	0.20	0.78	>50	50	>100	0.78	50
Methicillin-resistant <i>S. aureus</i> 69 ^c	0.20	1.56	>50	50	>100	1.56	50
<i>Candida albicans</i> IFO 1269	3.13	25	>50	50	>100	12.5	>50
<i>C. albicans</i> IFM 40009	6.25	25	>50	50	>100	12.5	>50
<i>Cryptococcus neoformans</i> TIMM 0354	0.39	3.13	25	12.5	50	1.56	12.5
<i>C. neoformans</i> TIMM 0362	≤ 0.20	3.13	25	6.25	25	1.56	6.25
<i>Aspergillus fumigatus</i> TIMM 0063 ^d	6.25	6.25	50	6.25	50	6.25	50
<i>A. fumigatus</i> IMF 4942 ^d	6.25	12.5	50	12.5	100	6.25	50

Test organism	MIC ($\mu\text{g/ml}$)						
	8	9	10	11	12	13	14
<i>Citrobacter freundii</i> IFO 12681	>50	>50	>12.5	>50	>50	100	>100
<i>Enterobacter cloacae</i> IFO 12935	>50	>50	>12.5	>50	>50	12.5	100
<i>Escherichia coli</i> NIHJ JC-2	>50	>50	>12.5	>50	>50	12.5	100
<i>Klebsiella pneumoniae</i> IFO 3317	25	50	>12.5	25	25	1.56	25
<i>Proteus vulgaris</i> GN 5298	50	>50	>12.5	50	50	3.13	50
<i>Pseudomonas aeruginosa</i> IFO 3445	>50	>50	>12.5	>50	>50	100	>100
<i>Serratia marcescens</i> 3759	>50	>50	>12.5	>50	>50	50	>100
<i>Streptococcus epidermidis</i> IFO 13889	50	50	12.5	6.25	12.5	0.78	6.25
<i>Enterococcus faecalis</i> IFO 12964	50	>50	12.5	6.25	50	0.78	6.25
<i>E. faecium</i> IFO 12367	25	50	6.25	1.56	50	0.39	6.25
<i>Staphylococcus aureus</i> IFO 12732	50	50	12.5	6.25	12.5	0.39	6.25
Methicillin-resistant <i>S. aureus</i> 4 ^c	25	50	6.25	3.13	12.5	0.20	6.25
Methicillin-resistant <i>S. aureus</i> 69 ^c	50	50	12.5	6.25	12.5	0.20	6.25
<i>Candida albicans</i> IFO 1269	50	50	12.5	6.25	50	6.25	12.5
<i>C. albicans</i> IFM 40009	50	25	12.5	6.25	50	6.25	25
<i>Cryptococcus neoformans</i> TIMM 0354	6.25	3.13	3.13	1.56	6.25	0.39	3.13
<i>C. neoformans</i> TIMM 0362	6.25	6.25	3.13	1.56	6.25	0.39	3.13
<i>Aspergillus fumigatus</i> TIMM 0063 ^d	25	6.25	12.5	6.25	6.25	3.13	12.5
<i>A. fumigatus</i> IMF 4942 ^d	25	6.25	12.5	6.25	12.5	6.25	12.5

^a MIC values were determined by an agar dilution method using mueller Hinton agar for antibacterial tests with incubation at 37°C for 18 hours and a Sabouraud Dextrose agar for antifungal tests with incubation at 30°C for 24 hours.

^b K salt.

^c Clinical isolate.

^d Incubation: 48 hours.

Table 2. ^1H NMR data of neopyrrolomycin analogs.

No	δ ppm
2	5.35 (2H, brs), 6.43 (2H, t, $J=2$ Hz), 6.70 (2H, t, $J=2$ Hz), 7.00 (1H, d, $J=2$ Hz), 7.10 (1H, d, $J=2$ Hz)
3	5.23 (1H, s), 6.44 (2H, t, $J=2$ Hz), 6.73 (2H, t, $J=2$ Hz), 6.95 (1H, dd, $J=8, 2$ Hz), 7.05 (1H, dd, $J=8, 2$ Hz), 7.25 (1H, t, $J=8$ Hz)
4	5.28 (1H, s), 6.40 (2H, t, $J=2$ Hz), 6.85 (2H, t, $J=2$ Hz), 6.97 (1H, d, $J=9$ Hz), 7.25 (2H, m)
5	5.27 (1H, s), 6.38 (2H, t, $J=2$ Hz), 6.83 (2H, t, $J=2$ Hz), 7.00 (2H, m), 7.27 (2H, m)
6	5.35 (1H, brs), 6.33 (2H, m), 6.60 (1H, m), 7.01 (1H, d, $J=2$ Hz), 7.10 (1H, d, $J=2$ Hz)
7	5.18 (1H, s), 6.37 (2H, m), 6.67 (1H, m), 7.00 (1H, dd, $J=8, 2$ Hz), 7.31 (1H, t, $J=8$ Hz)
8	5.15 (1H, s), 6.36 (1H, m), 6.63 (2H, m), 6.93 (1H, dd, $J=8, 2$ Hz), 7.03 (1H, dd, $J=8, 2$ Hz), 7.25 (1H, t, $J=8$ Hz)
9	5.22 (1H, s), 6.33 (1H, m), 6.77 (2H, m), 7.03 (2H, m), 7.23 (2H, m)
10	5.30 (1H, s), 6.28 (1H, d, $J=2$ Hz), 6.63 (1H, d, $J=2$ Hz), 6.98 (1H, dd, $J=8, 2$ Hz), 7.08 (1H, dd, $J=8, 2$ Hz), 7.33 (1H, t, $J=8$ Hz)
11	5.25 (1H, s), 6.23 (1H, d, $J=2$ Hz), 6.68 (1H, d, $J=2$ Hz), 6.98 (1H, d, $J=9$ Hz), 7.18 (1H, d, $J=2$ Hz), 7.30 (1H, dd, $J=9, 2$ Hz)
12	5.12 (1H, s), 6.25 (1H, d, $J=2$ Hz), 6.72 (1H, d, $J=2$ Hz), 7.07 (3H, m), 7.38 (1H, m)
13	5.68 (1H, brs), 6.38 (1H, d, $J=4$ Hz), 6.58 (1H, d, $J=4$ Hz), 7.02 (1H, d, $J=2$ Hz), 7.11 (1H, d, $J=2$ Hz)
14	5.45 (1H, s), 6.40 (1H, d, $J=4$ Hz), 6.63 (1H, d, $J=4$ Hz), 6.98 (1H, dd, $J=8, 2$ Hz), 7.08 (1H, dd, $J=8, 2$ Hz), 7.32 (1H, t, $J=8$ Hz)

drofuran gave the corresponding pyrroles **16a**~**16d**. **16a**: 64%, mp 86~90°C; **16b**: 90%, mp 80~82°C; **16c**: 75%, oil; **16d**: 50%, oil. De-*O*-methylation of **15a**~**15d** with AlCl_3 in benzene led to the neopyrrolomycin analogs **2**~**5**. **2**: 84%, mp 68~69°C; **3**: 91%, mp 89~92°C; **4**: 77%, oil; **5**: 86%, oil.

Selective chlorination of **16a**~**16d** with *N*-chlorosuccinimide (NCS) in DMF at room temperature provided **17a** and **17c** exclusively but mixtures of **17b** and **18b**, and **17d** and **18d**. **17a**: 56%, oil; **17b**: 55%, oil; **18b**: 13%, oil; **17c**: 68%, mp 53~55°C; **17d**: 43%, oil; **18d**: 5%, oil. De-*O*-methylation of **17a** and **17b** with AlCl_3 gave **6** and **7**, respectively. **6**: 33%, oil; **7**: 98%, mp 55°C (dec). Similar deprotection of **18b** and **18d** yielded **8** and **9**, respectively. **8**: 85%, mp 70~72°C; **9**: 78%, oil.

Further chlorination of **17b**~**17d** with NCS in DMF at 50°C followed by de-*O*-methylation produced the 2,4-dichloropyrrole derivatives **10**, **11** and **12** through the migration of a chlorine atom.^{3,4)} **10**: 40%, oil; **11**: 56%, mp 72~74°C; **12**: 50%, oil.

Regioselective bromination of **16a** and **16b** with *N*-bromosuccinimide in DMF at room temperature gave **20a** and **20b**. **20a**: 90%, oil; **20b**: 68%, oil. Lithiation with *n*-BuLi followed by treatment with CO_2 gas gave the carboxylic acids **21a** and **21b**, which were chlorinated by trichloroisocyanuric acid^{2~4)} to give **22a** and **22b**, respectively. **22a**: 83%, mp 215~217°C; **22b**: 70%, mp 205~208°C. These products were decarboxylated by heating with Cu

powder in quinoline, followed by de-*O*-methylation with AlCl_3 in benzene, to afford the neopyrrolomycin analogs **13** and **14**. **13**: 40%, oil; **14**: 70%, oil.

All analogs **2**~**14** showed antibacterial and/or antifungal activities as shown in Table 1. Remarkably, the simple 3,5-dichlorophenol analog **2** showed strong antibacterial activities against Gram-positive and -negative bacteria, but analogs **3**, **4** and **5** showed little activities. Also, the 3,5-dichlorophenol analogs **6** and **13** displayed stronger activities than the corresponding 3-chlorophenol analogs **7** and **14**. These findings suggest that the chlorine atom at the C-5 position of the benzene ring is essential for appearance of strong antibacterial activities.

The ^1H NMR (90 MHz, CDCl_3) of the target molecules (**2**~**14**) are summarized in Table 2.

Acknowledgments

We are grateful to Mochida Pharmaceutical Co., Ltd., Yamanouchi Pharmaceutical Co., Ltd. and Shikoku Chemicals Co. for the generous support of our program.

References

- 1) NOGAMI, T.; Y. SHIGIHARA, N. MATSUDA, Y. TAKAHASHI, H. NAGANAWA, H. NAKAMURA, M. HAMADA, Y. MURAOKA, T. TAKITA, Y. IITAKA & T. TAKEUCHI: Neopyrrolomycin, a new chlorinated phenylpyrrole antibiotic. *J. Antibiotics* 43: 1192~1194, 1990
- 2) TATSUTA, K. & M. ITOH: Total synthesis of chlo-

- minated phenylpyrrole antibiotics, (+)- and (-)-neopyrrolomycins. *Tetrahedron Lett.* 34: 8443~8444, 1993
- 3) TATSUTA, K. & M. ITOH: Synthesis and biological evaluation of neopyrrolomycin analogs. *J. Antibiotics* 47: 262~265, 1994
- 4) TATSUTA, K. & M. ITOH: Total synthesis of (+)- and (-)-neopyrrolomycins, chlorinated phenylpyrrole antibiotics. *Bull. Chem. Soc. Jpn.*, in press.
- 5) KIMURA, T.; N. WATANABE, M. MATSUI, K. HAYASHI, H. TANAKA, I. OHTSUKA, T. SAEKI, M. KOGUSHI, H. KABAYASHI, K. AKASAKA, Y. YAMAGISHI, I. SAITOU & I. YAMATSU: Structure-activity relationship of a series of phenylureas linked to 4-phenylimidazole. Novel potent inhibitors of acyl-CoA: Cholesterol O-acyltransferase with antiatherosclerotic activity. 2. *J. Med. Chem.* 36: 1641~1653, 1993